

# Speak from “Synthetic Biology: Licensing Bacteria to Kill”

*Andrew Jermy:*

Pyocin S5-producing *E. coli* inhibited the growth of *P. aeruginosa* by >99%.

*Nature Reviews Microbiology* | AOP, published online 12 September 2011; doi:10.1038/nrmicro2660

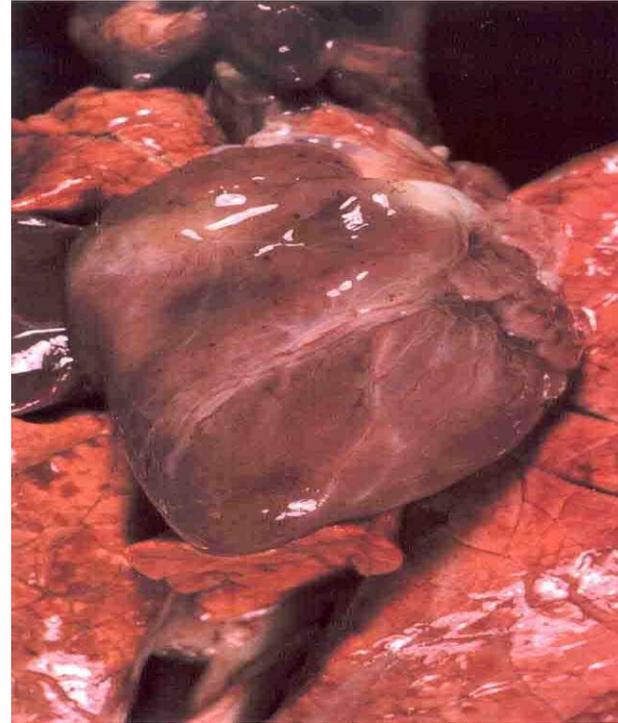
Speaker: Zhi-Ming Xie

# Background

Diseases caused by *Pseudomonas aeruginosa* (PA)



G<sup>-</sup>



septicaemia

# Several Therapy Strategies

- Antibiotics(one or more species)  
Multi-drug resistance
- Phage therapy: Because of potential employing virus in bacterial infection ,it is limited .
- Development novel ,unconditional antimicrobial strategies is immediately required.

# Bacteriocins—Pyocin Type S5

- Ribosomally synthesized antibacterial peptides.
- Specific and effective against closely related species
- New candidate of the next generation antibacterial agent
- Pyocins produced by PA are narrow-spectrum bacteriocins and classified into 3 types(R,F,S).
- S5 is soluble and consists two parts:the larger can kill while the smaller is prevent the PA from killing (immunity protein).

# Quorum sensing

- An organic chemical signaling cascade responded to different physiological activities(motility)
- PA can produce AHL 3O-C<sub>12</sub>HSL as the signal.
- Infected Extracellular concentration of AHL 3O-C<sub>12</sub>HSL is 1.0E-6 to 1.0E-4M.

# Goal—Method



Hunter  
(Detect AHLs)

Detect goal

the Sensing Device



Trigger

Fire

the Lysis Device (E7 Protein)



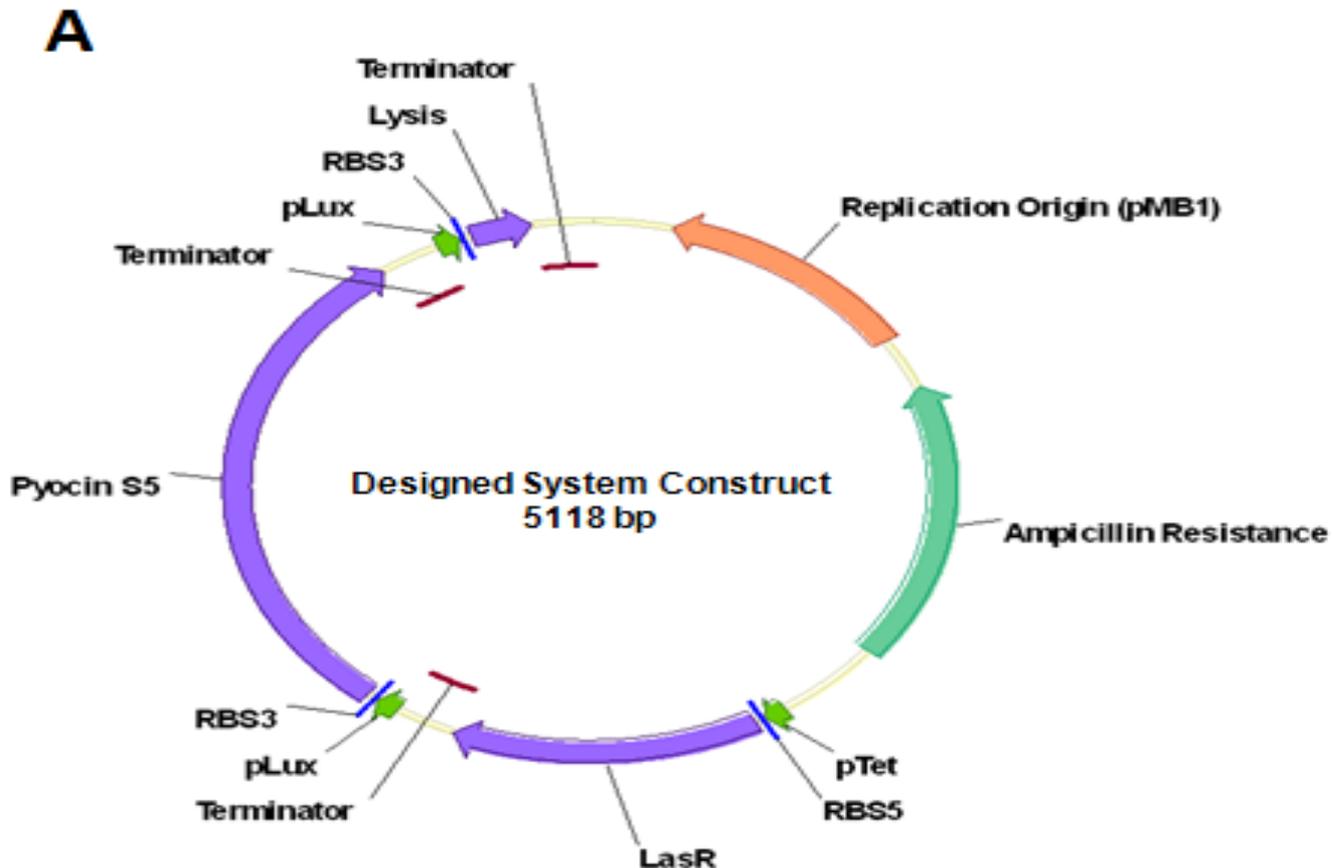
Bullet

kill PA

the Killing Device (Pycoin S5)

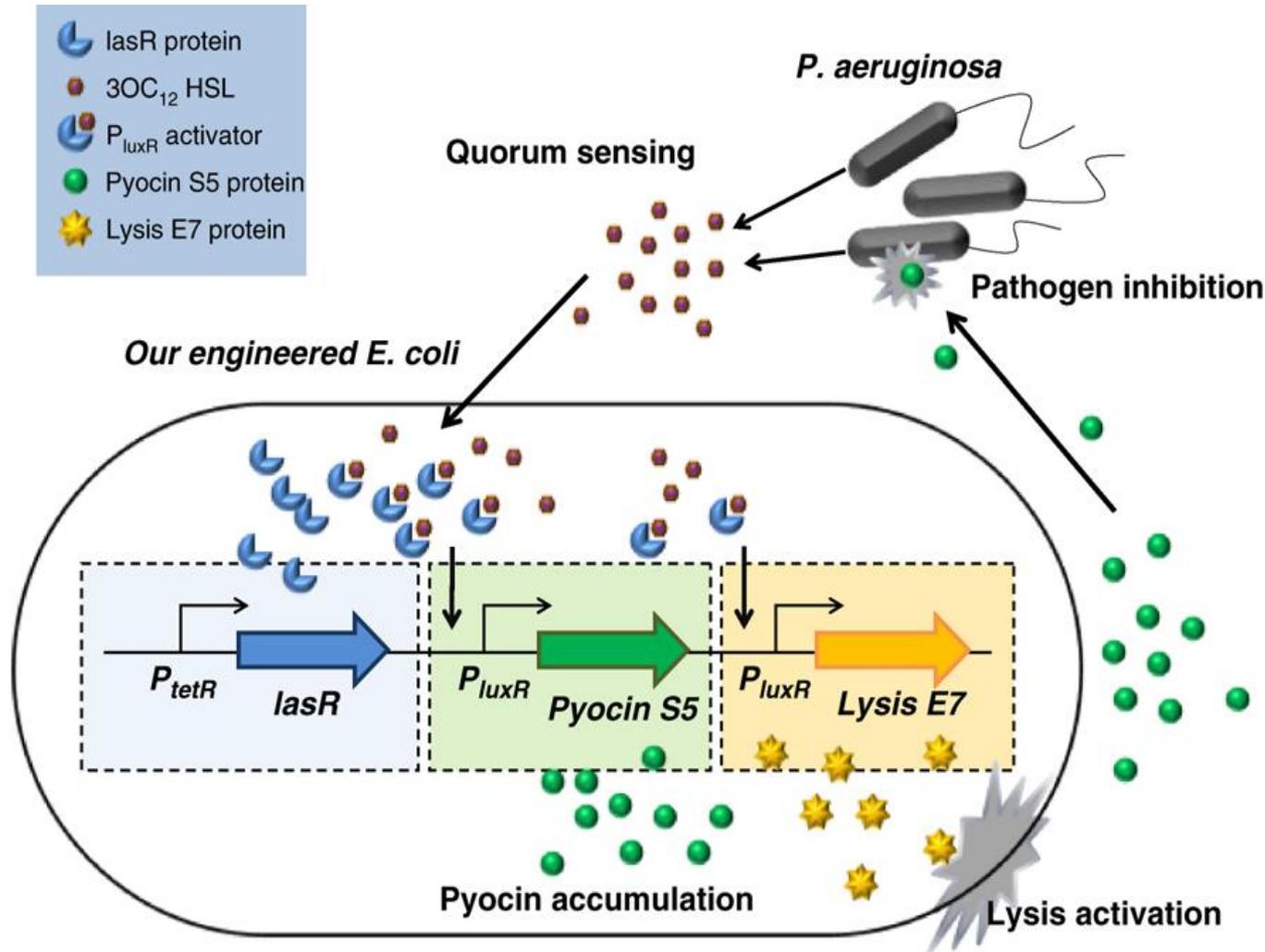
# Methods 、 Results and Discussion

## Design of the sensing and killing genetic system



The final engineered system, pTetR-LasR-pLuxR-S5-pLuxR-E7. The system recognizes input chemical signals from *P. aeruginosa* and produces S5 pyocin and E7 lysis proteins.

# Continued .....



1、3OC<sub>12</sub>HSL produced by PA caused quorum sensing

2、3OC<sub>12</sub>HSL can bind tetR expressed constitutively

3、3OC<sub>12</sub>HSL bind tetR expressed constitutively

# Characterization and model of the sensing device (pTetR-LasR-pLuxR-GFP)

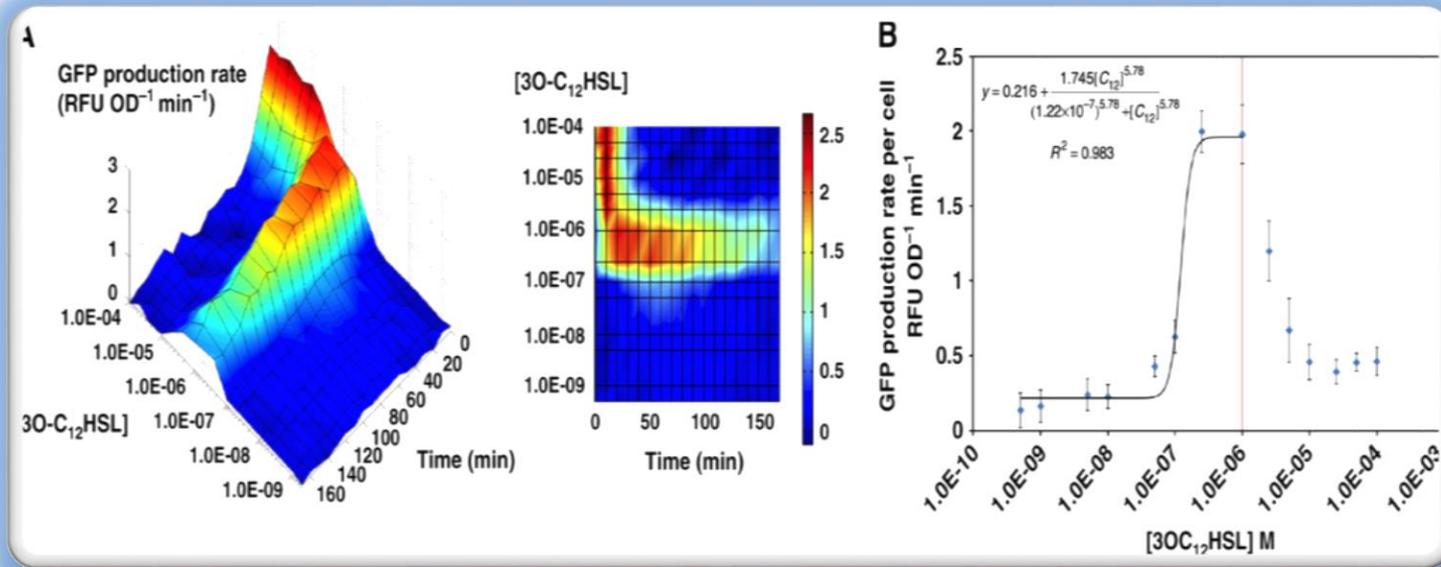


Fig.A: 1.  $3O-C_{12}HSL > 1.0E-7M$  , GFP sharply increase;  
 2.  $3O-C_{12}HSL < 1.0E-5$  , GFP sharply decline.

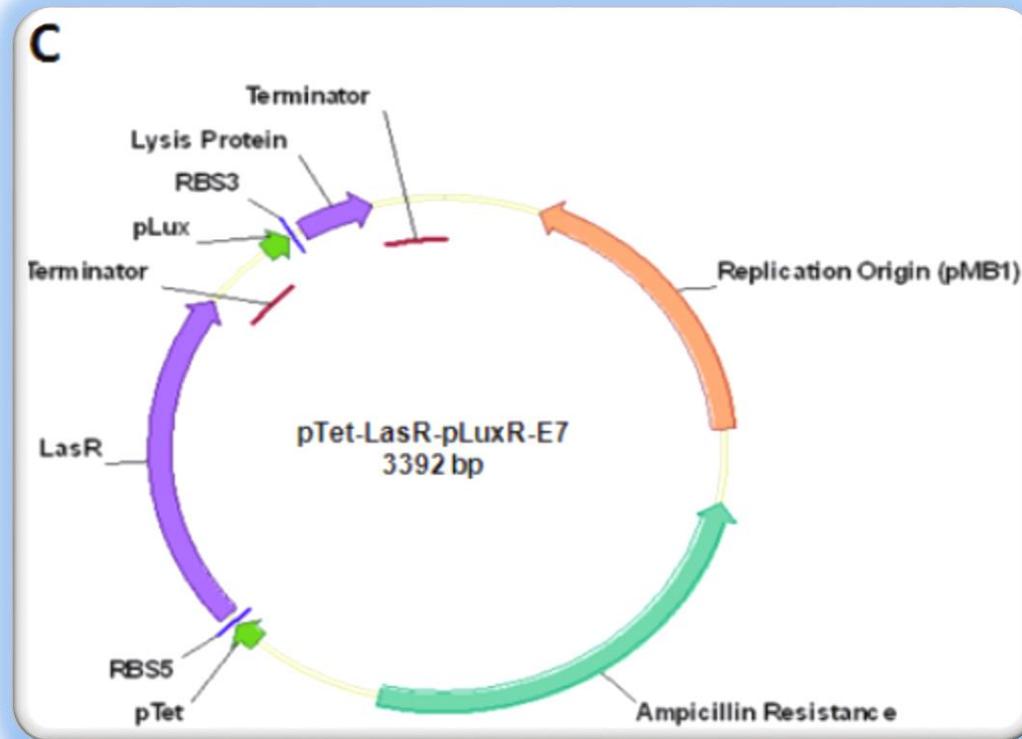
Fig.B: 1.The static relationship between input( $3O-C_{12}HSL$ )and output(GFP) is Hill equation(input  $< 1.0E-6M$ ).  
 2.  $1.0E-6M < 3O-C_{12}HSL < 3.3E-7M$  , the output is the maximum(1.96RFU/OD)  
 3. Half-maximum:  $1.2E-7M$

Summary: The designed sensing device is sensitive enough to detect natively produced by PA( $1.0E-6$  to  $1.0E-4$ )

## Detection of the native autoinducer produced by *P. aeruginosa* (pTetR-LasR-pLuxR-GFP)

- pTetR-LasR-pLuxR-GFP is co-cultured with *P. aeruginosa* ln7 (ln7 is sensitive to pyocin S5)
- Result: ln7 was 1.375 RFU/OD, between the minimum synthesis rate and the half-maximal of the sensing device.
- Sum: The sensing device can detect the pyocin S5 natively produced by PA.

# Characterization of the lysing device (pTetR-LasR-pLuxR-E7)



Methodology: PA and lysis device co-cultured and then induced by 0 to  $1.0 \times 10^{-8} \text{M}$   $3\text{OC}_{12}\text{HSL}$ . Subsequently, measured the  $\text{OD}_{600}$  and viewed the cell morphology by FESEM(场发射扫描电子显微镜).

# Continued .....

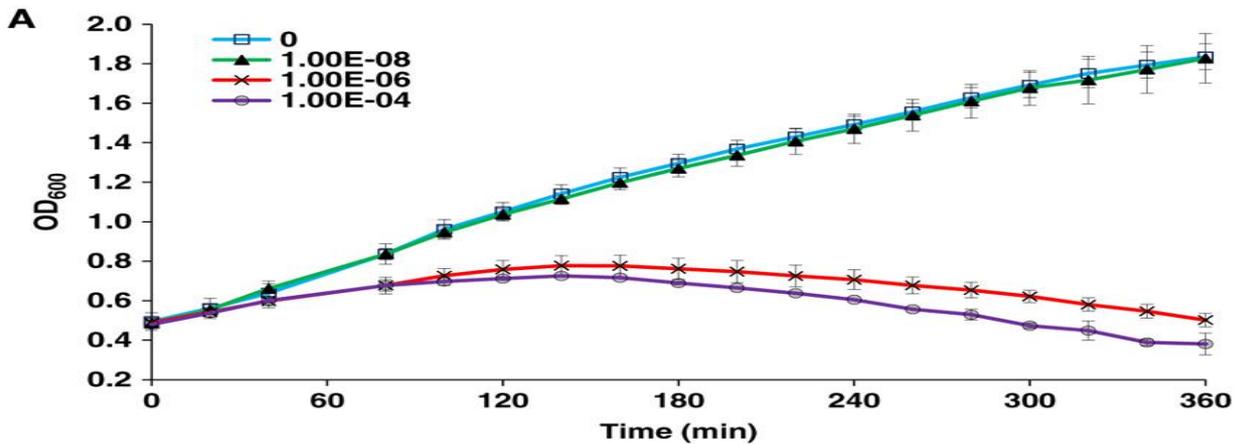


Fig. A: 3OC<sub>12</sub>HSL were 1.0E-6M and 1.0E-4M exhibit the cell growth

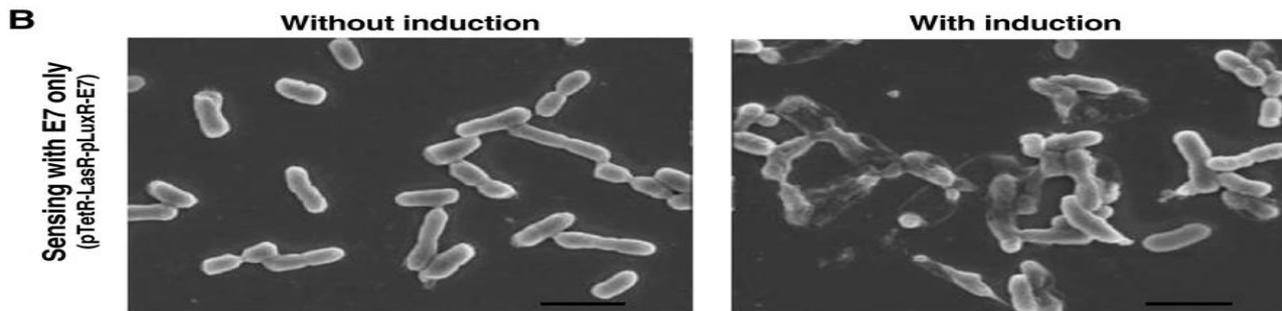


Fig. B: The cell induced by 3OC<sub>12</sub>HSL appeared shriveled with corrugated

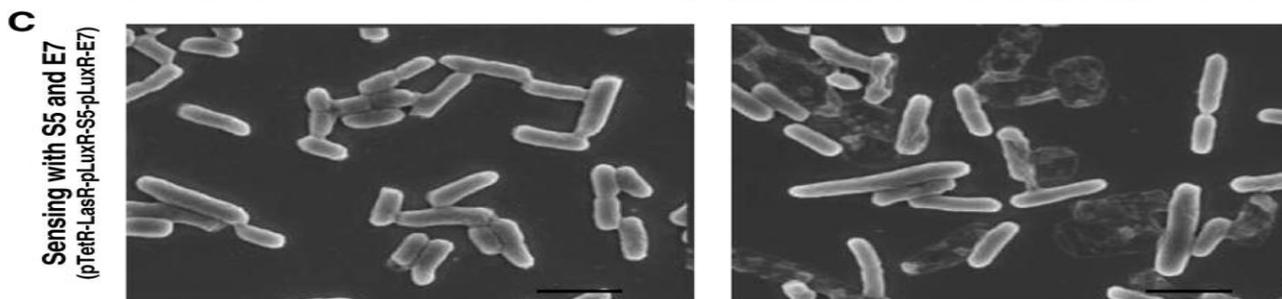
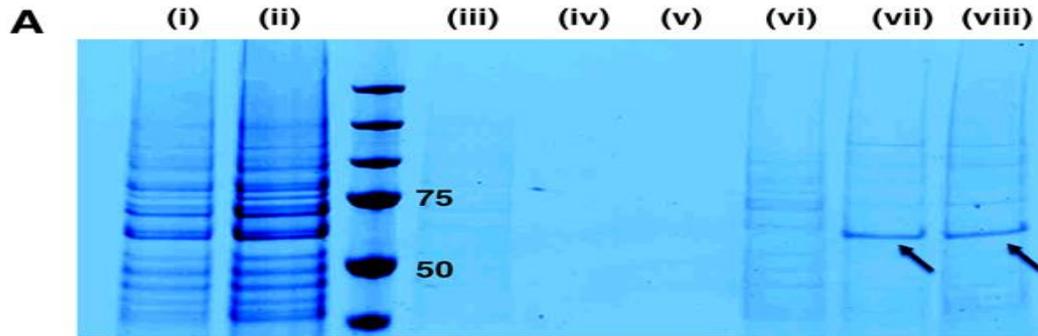
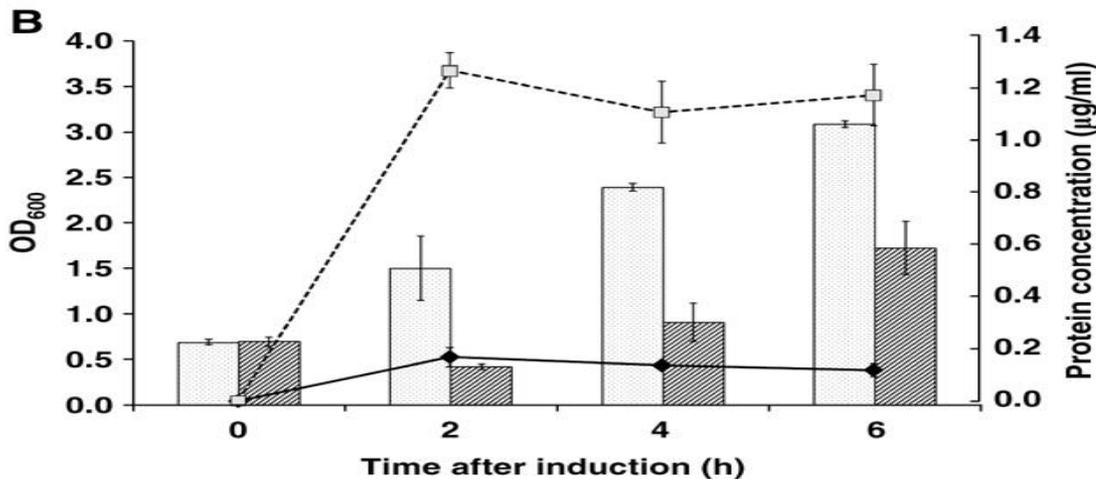


Fig. C: PA and the final system co-cultured. The cell induced by 3OC<sub>12</sub>HSL appeared shriveled with corrugated.

# Continued.....



i, ii S5、s5+E7总蛋白；iii-v: S5 诱导0.2.4h后  
vi-viii: S5+E7诱导0.2.4h后  
只有S5+E7在诱导2、4h后表达出S5  
(57kD)



1、impulse release of pyocin S5 at 2 h after induction, followed by a sustained steady-state release in the final system (dotted lines).

2、OD of the final system was characterized by an initial decrease at 2 h after induction, indicative of the onset of lysis, after which the regrowth of engineered E. coli occurs (shaded bar).

□ Cell growth of pTetR-LasR-pLuxR-S5      —●— His-tagged S5 of pTetR-LasR-pLuxR-S5  
 ■ Cell growth of pTetR-LasR-pLuxR-S5-pLuxR-E7      -□- His-tagged S5 of pTetR-LasR-pLuxR-S5-pLuxR-E7

Line: Pyocin S5      Bar graph: OD<sub>600</sub>

# Verification of the final system with the sensing, killing, and lysing devices

Fig. A: Only the designed microbe can inhibit PA.

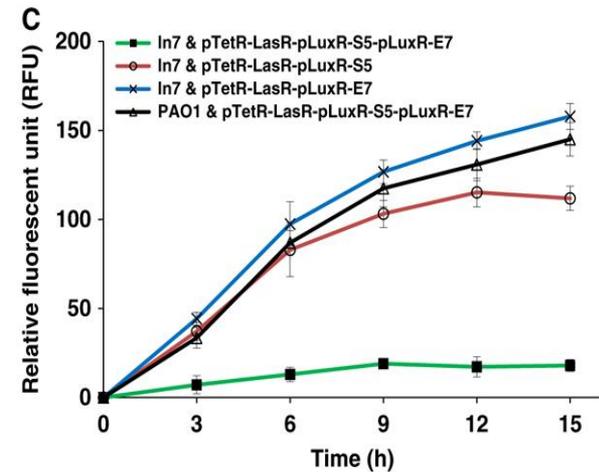
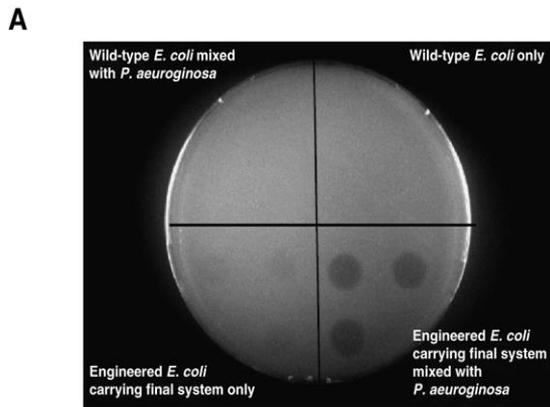


Fig. C: Only the designed microbe can inhibit PA.

Fig. B: LIVE/DEAD Stain.  
Dead---PI stained(Red)  
Live---SYTO9 stained(Green)

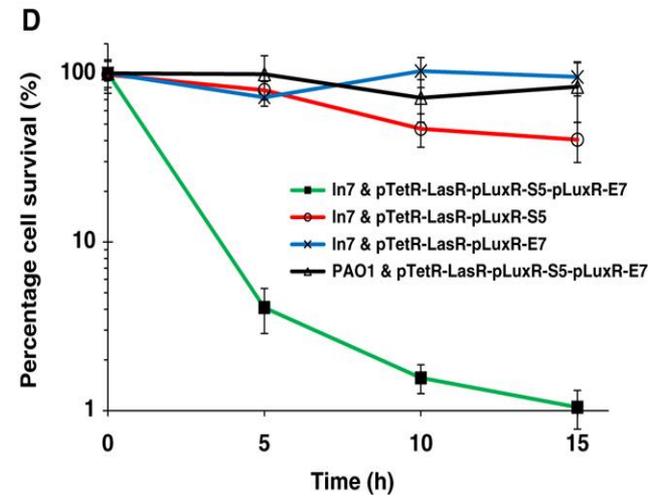
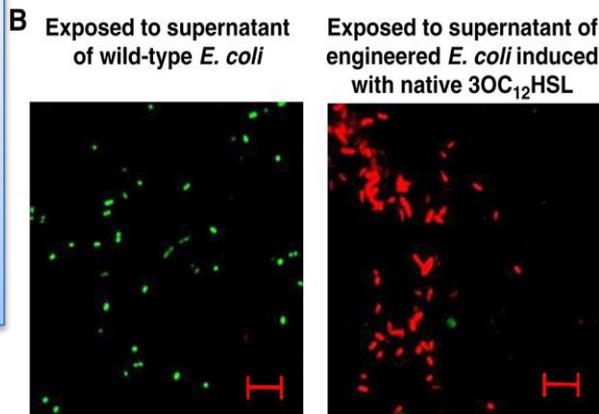


Fig. D: Only the designed microbe can inhibit PA. (about >99%)

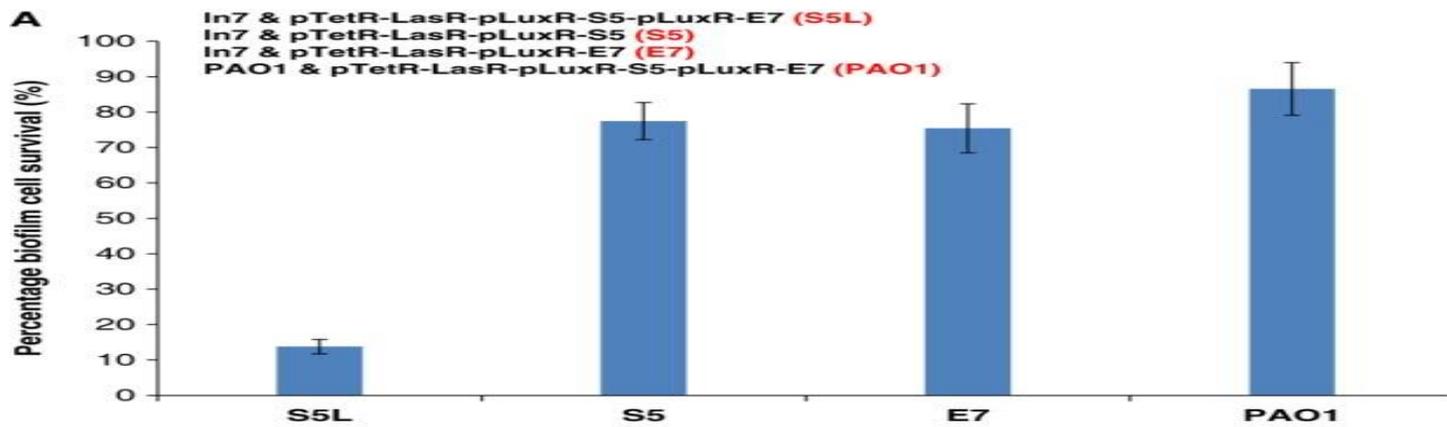
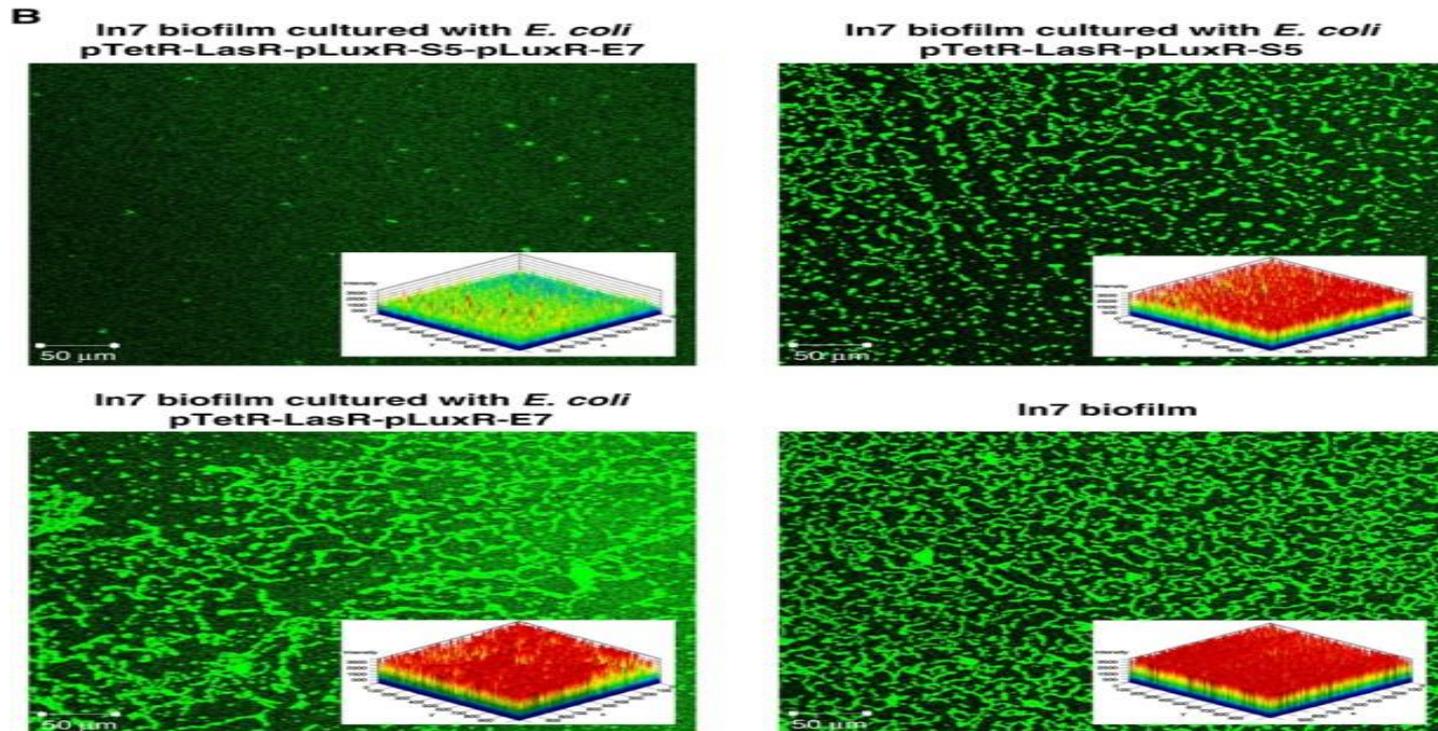


Fig. A:  
inhibition efficiency is close to 90%.



# Conclusions

- 1、 The engineered *E.coli* can inhibit biofilm formation during the initial attachment phase and prevent subsequent progression into mature microcolonies ( $P_{15}$ ) .
- 2、 The engineered *E.coli* can effectively inhibitor or kill PA.

Thank you for your patience !

Now, Let's Communicate !