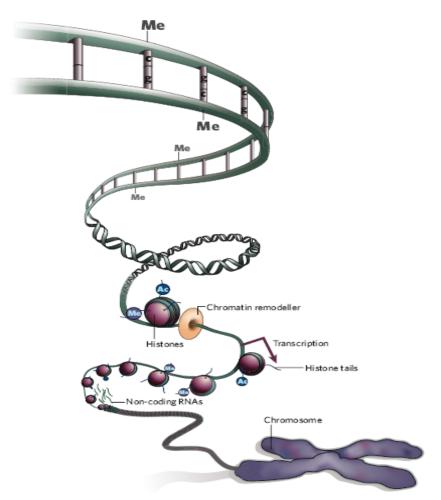
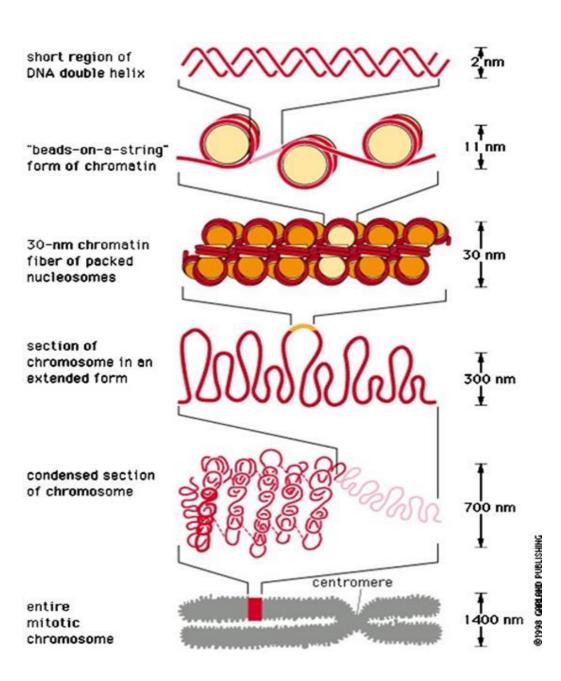
Complexity of chromatin folding is captured by the strings and binders switch model



王文韬 周群丰





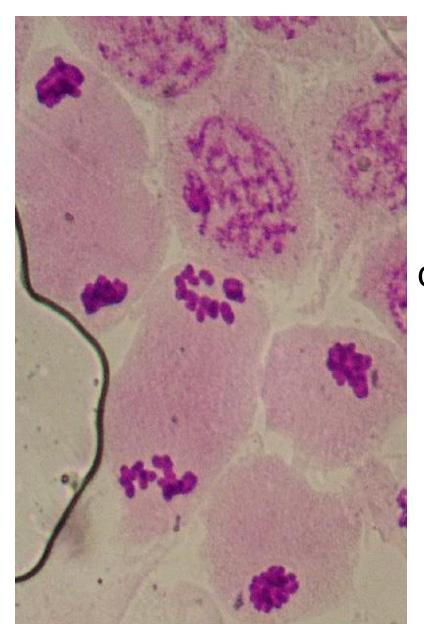
chromosome compaction

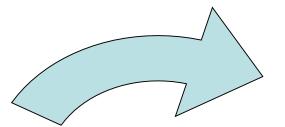
Chromosome compaction

mitotic cycle dynamic compaction Hetero- and eu-chromosome Compaction (stable territories)

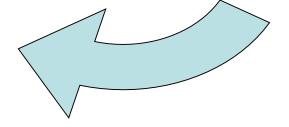
Interphase dynamic compaction (discrete territories)

mitotic cycle dynamic compaction

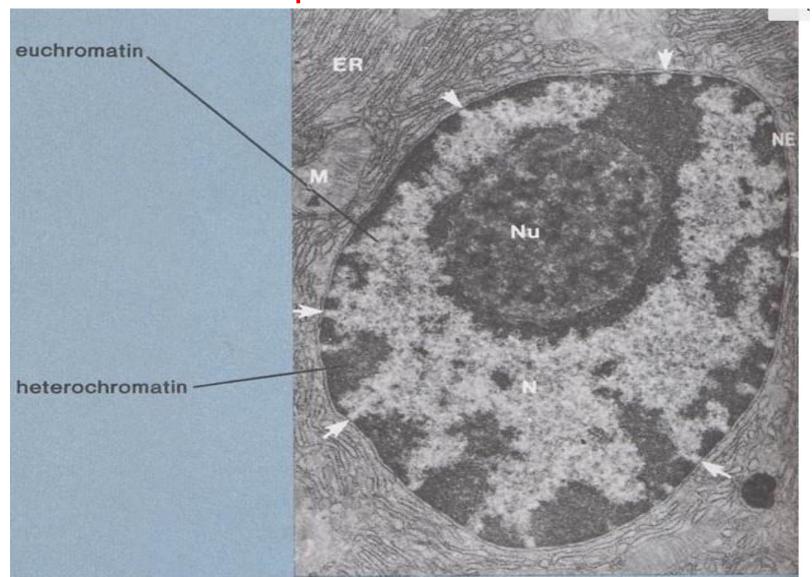


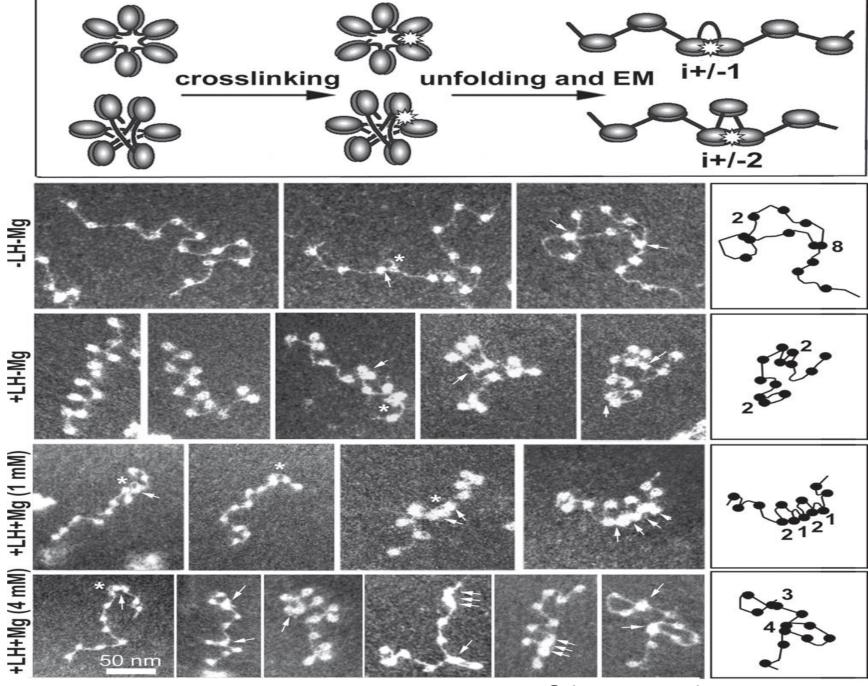


compaction unfolding



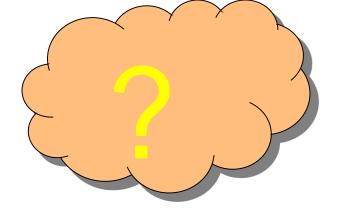
Hetero-chromomatin and euchromamtin ----stable compaction





Grigoryev et al.2009

•Spatial genome organization is guided by intra-and inter-chromosomal interactions mediated by nuclear components that include transcription factors, transcription and replication factories, Polycomb bodies, and contacts with the lamina.



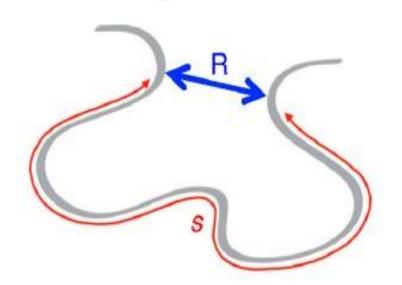
How binding of diffusible factors to specific genomic regions drives chromatin folding remains poorly understood.

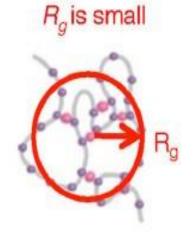
Three key tech. to analysis compaction

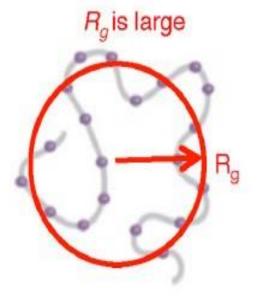
FISH: Imaging of single loci
3C (chromosome conformation
capture approaches): genome-wide
mapping of chromatin interactions
Hi-C(A global analysis of genomewide 3C)

Mean spatial distance

Gyration Radius

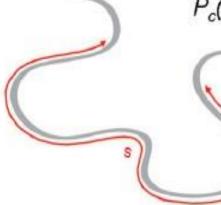


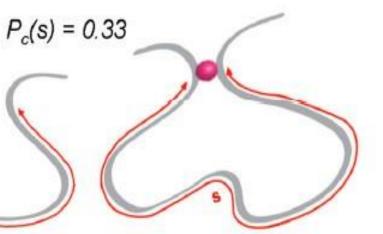




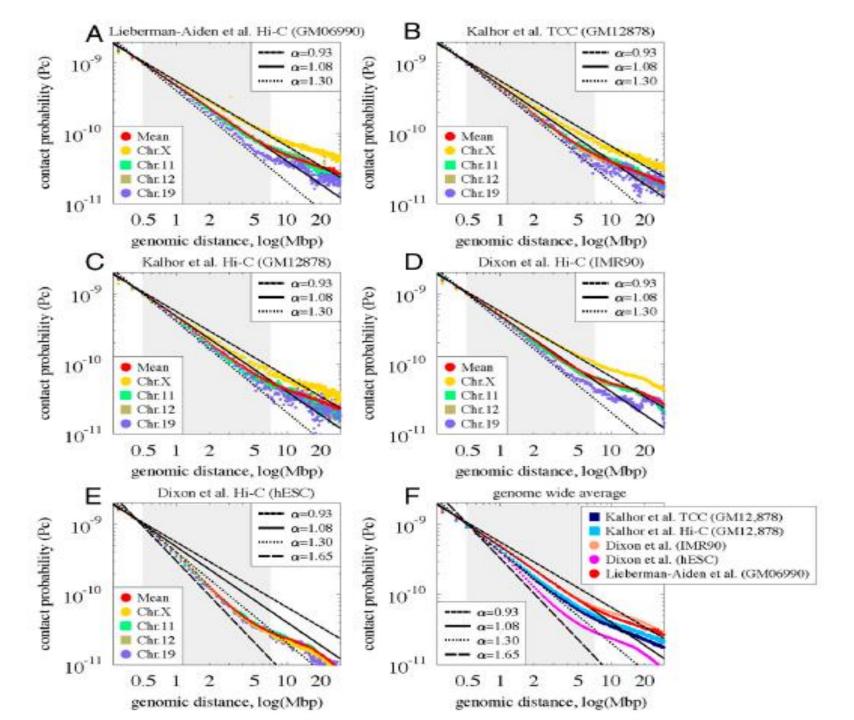
Contact Probability







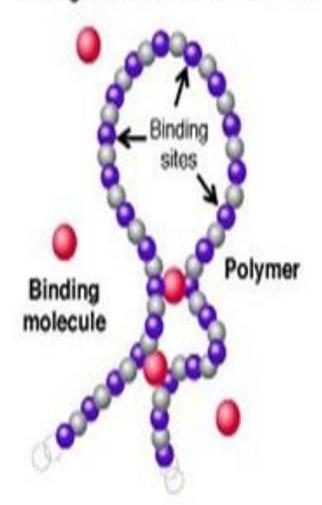
Cellular material	Technique	Genomic region analysed (resolution)	Mean square spatial distance, $R^2(s)$, as a function of genomic separation, s	Contact probability, $P_c(s)$, as a function of genomic separation, s	Ref.
Human female fibroblast cells	3D-FISH	Chr1q, 27Mb (~1Mb) and Chr11q, 75Mb (~3Mb)	s = 0.4-2Mb, $R^2(s)$ increases (v = 0.33) s >10Mb, $R^2(s)$ reaches a plateau (v = 0)	N/A	9
Mouse pre-pro-B cells (E2A+) and pro- B cells (RAG+)	3D-FISH	Igh locus, 3Mb (~300kb)	s <0.5Mb, $R^2(s)$ increases (v = 0.25 for pre- pro-B cells, v = 0.1 for pro-B cells) s >0.5Mb, $R^2(s)$ reaches a plateau (v = 0; both cell types)	N/A	10
Mouse NIH-3T3 fibroblasts	3D-FISH	Chr14, 4.3Mb (200kb)	$s < 3.5$ Mb, $R^2(s)$ increases (v ~ 0.5) $s > 3.5$, $R^2(s)$ may plateau	N/A	11
Human male fibroblast cells	2D-FISH	Chr4 (10Mb)	s <50Mb, $R^2(s)$ increases ($v \sim 0.5$) s >50Mb, $R^2(s)$ increases ($v \sim 0.3$)	N/A	12
Human lymphoblastoid cell line	Hi-C	Genome-wide (1Mb)	N/A	s = 0.5-7Mb, $P_c(s)$ decreases approximately as a power law, with exponent α = 1.08	13
Drosophila embryos	Simplified Hi-C	Genome-wide and Repressive epigenetic classes	N/A	Genome wide: contact frequencies decrease approximately as a power law, with exponent α = 0.85 Repressive epigenetic classes: contact frequencies decrease approximately as a power law, with exponent α = 0.70	14



SBS

- In "strings and binders switch" (\$
 conformations are established t
 factors (binders) to binding sites
 give rise to a variety of stable cl
 coexist in the nucleus. Chromati
 changes in binding site distribinding affinity, in a switch-like
 values via thermodynamics
- More importantly, SBS moderal current experimental data on FISH, Hi-C and 3C approaches

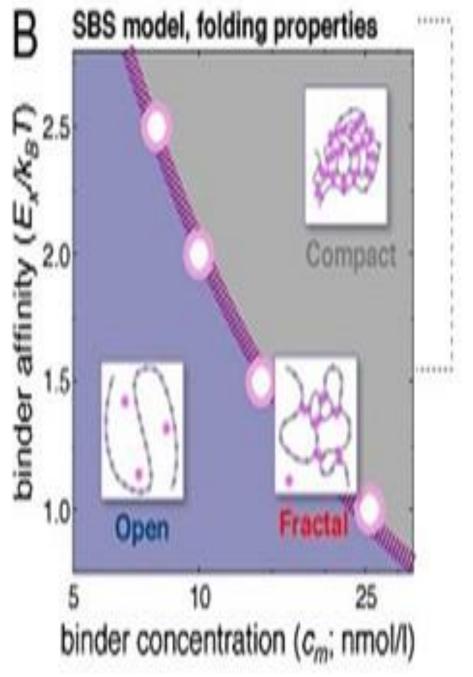
Strings & Binders Switch model



- SBS mode evaluated the dyr the polymer using extensive known range of the biochemi binders) and E_x (binders affi
- E_X=2K_BT (kB is the Boltzma
- in kelvin)

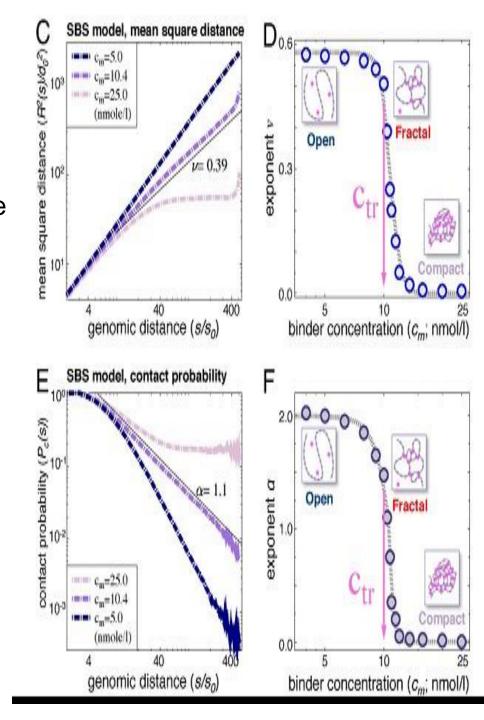
 E_x change with the C_m

• C_{tr} (threshold concentration)

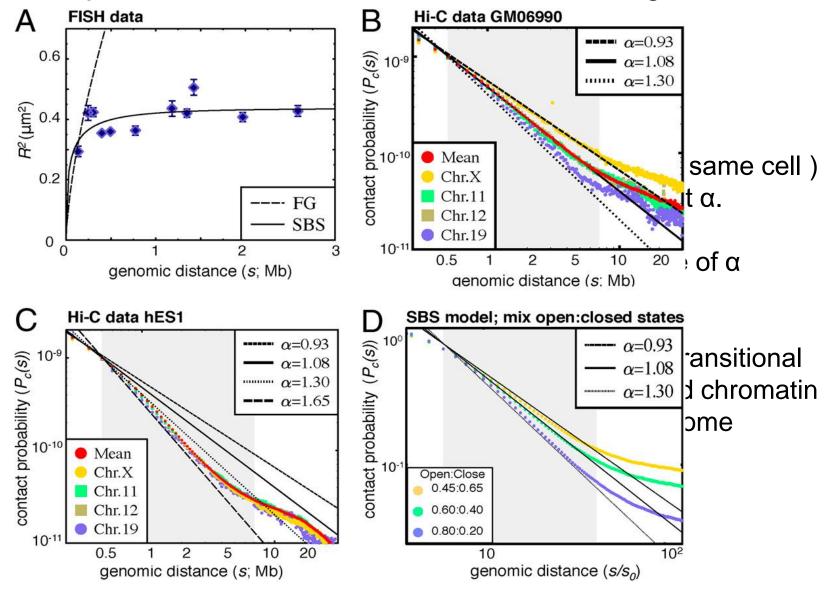


In the SBS model, the shape of the two functions $R_2(s)$ and Pc(s) is sensitive to the concentration of binding molecules, C_m $Pc(s)\sim 1/S\alpha$

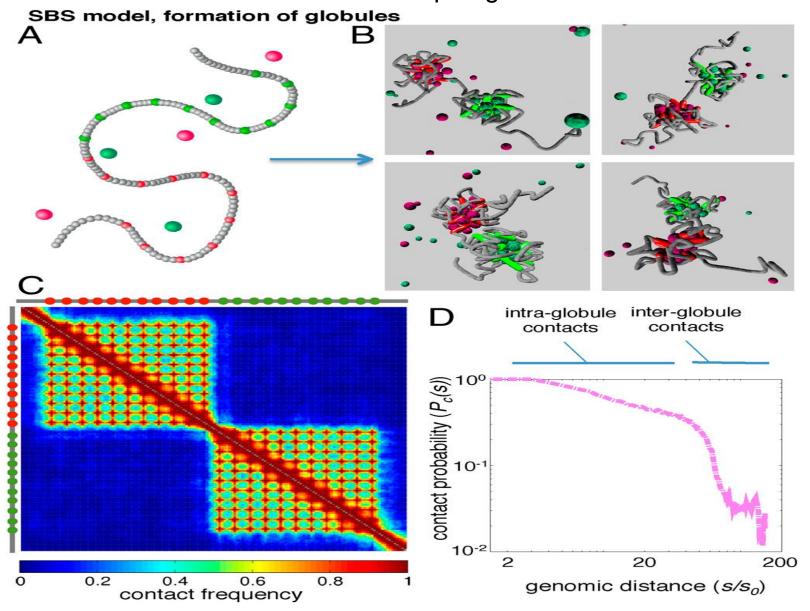
 $C_m \& C_{tr}$ $R^2(s) \sim S_{2v}$



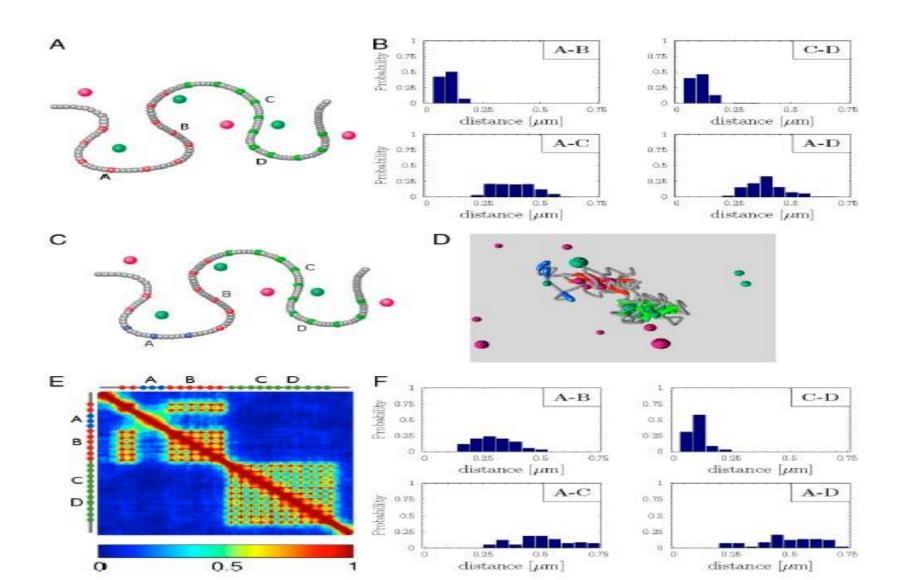
Comparison of the SBS and Other Models Against



The SBS Model Reproduces the Organization of Chromatin in Topological Domains.



changed the state of three contiguous sites from binding the red binders to no longer having affinity to binders



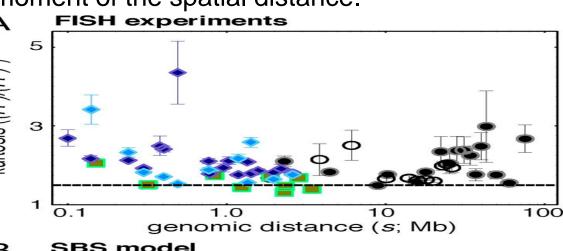
The SBS Model Reproduces the Dynamic Folding Behaviors of Chromatin.

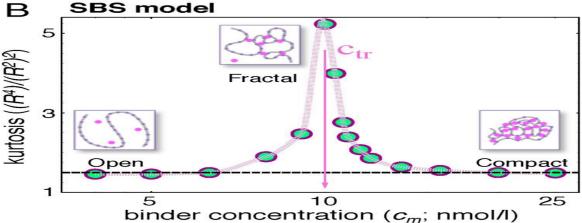
• As a final test to the power of the SBS model, researcher investigated the kurtosis K =(R⁴(s))/(R²(s))², which is the ratio of the fourth and second moment of the spatial distance.

K carry information of genomic locus and cell – cell variation, and dependent on Cm

K=1.5, corresponding is compact and open

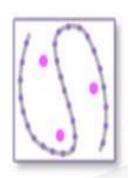
K=5,corresponding region is transition



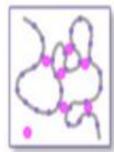


Discussion

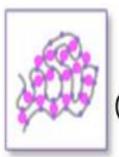
- 1. Interphase nuclei exhibit dynamic chromatin structures that change in response to cellular signals and influence patterns of gene expression.
 2. genomic architectures sharp regulation of nuclear architecture can be obtained reliably by simple strategies, such as protein up-regulation or modification, without the need to fine tune these specific parameters.
- 3. SBS mode analysis strongly supports the conclusion that the principles of chromatin folding in interphase nuclei cannot be recapitulated by a single "universal" conformational state (and its given α).
 - 4. SBS model illustrates key physical concepts and basic required ingredients to explain chromatin folding in a variety of states identified in living systems, a variety of specific binding factors exist, where different regions can spontaneously fold into different chromatin states
- 5. polymer scaling theory ensures that the exponents in R² and Pc are independent of the minute details of the system considered and reflect universal properties
- 6The thermodynamic mechanisms discussed, which are robust and independent of specific molecular details, will be relevant to many cellular and nuclear processes requiring spatial organization
 7.SBS mode is a powerful mode to analysis chromatin structure organization.



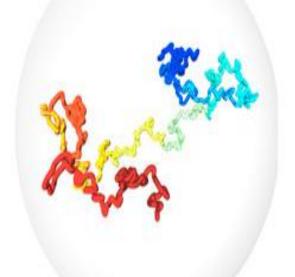
Open Conformation



Fractal Conformation



Compact Conformation

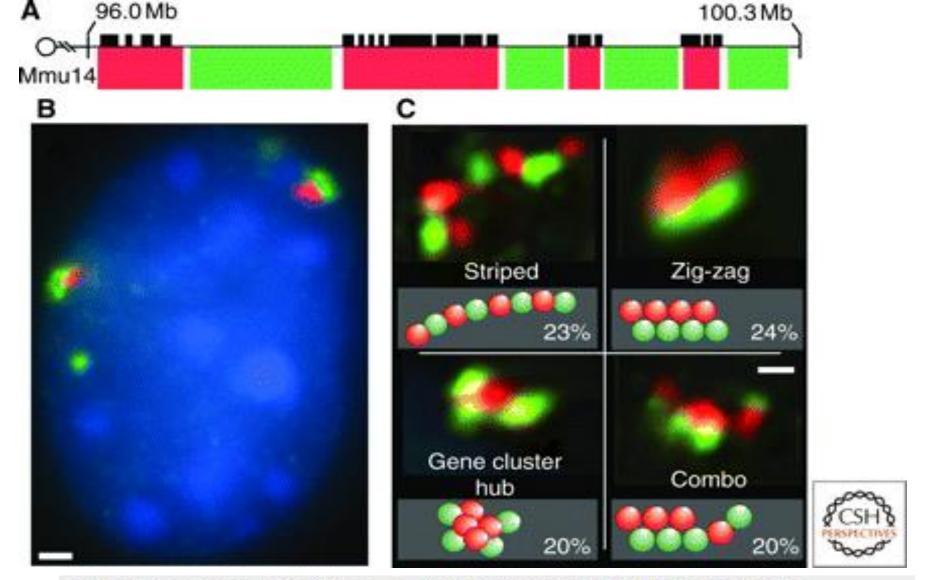






threshold concentration (C_{tr})

Binder concentration (c_m)



(A) A large locus consisting of \sim 4-Mbp region containing regions of gene "deserts" (red fluorescence) and gene clusters (green fluorescence) is seen in the nucleus in multiple configurations (C). In general, gene deserts are more closely associated with the heterochromatin at the nuclear periphery (B). Scale is 1 μ m. From Shopland et al. 2006.

Thanks for your attention